

An investigation of the toxic effects of a herbal formulation with anti-carcinogenic properties

*S. S. Iddamaldeniya¹, S. M. D. N. Wickramasinghe¹, M. I. Thabrew², N. Ratnatunge³,
M. G. Thammitiyagodage⁴

The Ceylon Journal of Medical Science 2005; 48: 13-23

Abstract

A decoction prepared from a mixture of *Nigella sativa* seeds, *Hemidesmus indicus* root, and *Smilax glabra* rhizome used by some traditional medical practitioners in Sri Lanka is considered to be useful for the treatment of cancer patients. However, there is a lack of information about any adverse effects of this decoction. Experiments were carried out using Wistar rats and ICR mice as the experimental model, to evaluate any adverse effects mediated by the above decoction.

Results of the investigations showed that administration of the decoction (at doses of 4g/kg body weight/day and 6g/kg body weight/day) to rats for three months had no adverse effects on the liver functions (as assessed by its effects on serum levels of alanine and aspartate aminotransferase and alkaline phosphatase) or haematological parameters (red blood cell count, white blood cell count, haemoglobin concentration, packed cell volume, mean corpuscular haemoglobin, mean corpuscular volume and mean corpuscular haemoglobin concentration). No significant pathological changes were observed in sections of the major body organs (liver, heart, lungs,

stomach, duodenum and kidney) of animals treated with the decoction for three months.

The investigations also demonstrated that the decoction did not have anti-ovulatory, anti-implantation, spermicidal activity. An attempt to determine the LD₅₀ concentration was unsuccessful. Even at a dose equivalent to 40 times (240g/kg/day) the normal therapeutic dose (6g/kg/day), no mortality or other toxic symptoms (loss of consciousness, salivation, muscle tremor, incoordination, hyperaesthesia, polyuria, anuria, polydipsia, defecation, piloerection, changes in locomotor activity, changes in posture, ataxia and loss of reflexes) were observed. Three month treatment with the decoction also did not produce any changes in average feed consumption, average body weight: liver weight ratios, or the general behaviour of the animals.

Key words: Rats; Ayurveda decoction, hepatotoxicity, reproduction parameters, anti-carcinogenic.

Introduction

A decoction prepared from a mixture of *Nigella sativa* Linn (Family: Ranunculaceae,

1. Department of Biochemistry, Faculty of Medical Sciences, University of Sri Jayawardenepura, Gangodwila, Nugegoda.
2. Department of Biochemistry and Clinical Chemistry, Faculty of Medicine, University of Kelaniya, Ragama.
3. Department of Pathology, Faculty of Medicine, University of Peradeniya, Peradeniya.
4. Animal Centre, Medical Research Institute, Colombo 8.

* Author for correspondence E-mail: drsamsi2003@yahoo.com

S: Kaluduru, T: Karungiragam) seeds, *Hemidesmus indicus* (L) R.Br. (Family: Asclepiadaceae, S: Iramusu, T: Arakkan) root, and *Smilax glabra* Rox B. (Family: Liliaceae, S: Cheena Ala) rhizome is used by some traditional medical practitioners in Sri Lanka for treatment of cancer (personal communication, Ayurveda Dr. N. Jayathilake, Physician, Bandaranaike Memorial Ayurveda Research Institute (BMARI), Navinna, Maharagama). Investigations with Wistar rats have shown that the decoction can significantly protect against diethylnitrosamine-induced hepatocarcinogenesis (1).

The value of any anti-carcinogenic agent would depend not only on its therapeutic efficacy but also on its lack of toxicity. Any anti-cancer drug with therapeutic value would have to be administered over a relatively long period. It must therefore be free of acute and chronic adverse effects. Therefore, a study was undertaken in rats, to evaluate any possible toxic effects mediated by long term (3 months) administration of the decoction comprising of *Nigella sativa* seeds, *Hemidesmus indicus* root, and *Smilax glabra* rhizome.

Material and Methods

Experimental animals

In all experiments Wistar rats (8 week old littermates, $190 \pm 10\text{g}$) and ICR mice ($20 \pm 5\text{g}$) were used and maintained in a temperature-controlled room ($25^\circ\text{C} \pm 2^\circ\text{C}$) under 12 hours light/dark cycle (dark phase 6 p.m. to 6 a.m.). They were fed with a standard laboratory diet containing 19% crude proteins, 3.8% fiber and 4400 kcal of energy, prepared by the Medical Research Institute, Sri Lanka, based on a formula recommended by the WHO, and water *ad libitum* (2).

Plant material

Dried rhizome of *Smilax glabra*, dried seed of *Nigella sativa* and dried root of *Hemidesmus indicus* were purchased locally, and identities were confirmed by Mr. Gunarathne Silva, Botanist, BMARI, Navinna, Maharagama, Sri Lanka. Voucher samples were placed at the BMARI.

Preparation of the decoction

The plant decoction was prepared according to the method recommended by traditional medical practitioners for the administration to cancer patients (personal communication, Ayurveda Dr. N. Jayathilake). Twenty grams each of *Nigella sativa* (dried seeds), *Hemidesmus indicus* (dried root, cut into small pieces) and *Smilax glabra* (dried rhizome, cut into small pieces) were mixed and boiled in 1.6 litres of distilled water and final volume was reduced to 200ml by boiling over 3 hours.

Dosage and administration of decoction

The decoction was administered to rats using a Sondi needle by gastric gavage method. The effect of two doses of the decoction was studied. Dose 1 was 4g/kg body weight/day. This dose corresponds to the normal therapeutic dose administered to adult humans as calculated based on the relative surface areas of human and rat. Dose 2 provided 6g decoction/kg body weight/day.

Yield of the decoction was 0.33g/ml. Calculation of the volume of decoction that should be given was done according to the required dose of the decoction and the body weights of rats and mice ($190 \pm 10\text{g}$ and $20 \pm 5\text{g}$ respectively). Whenever, the calculated volume exceeded the maximum volume that could be given to a rat or mouse (5ml/rat or 1ml/mouse), the calculated volume was heated over burner and reduced down.

Determination of LD₅₀

To determine the dose of the decoction that would cause the death of 50% of the test animals (LD₅₀), 20 female and 20 male ICR mice were randomly divided into four groups of ten each (5 females and 5 males). Groups 1 to 4 were orally fed with a single dose of the decoction 6g/kg body weight/day, 60g/kg body weight/day, 120g/kg body weight/day and 240g/kg body weight/day respectively. After single dosing with different concentrations of the decoction, mice were observed for seven days for any mortality, loss of consciousness, hyperaesthesia, salivation, muscle tremor and incoordination, polyuria, anuria, polydipsia, defecation, piloerection, and changes in posture, ataxia and loss of reflexes.

Effects on haematological parameters

Male Wistar rats (n=18, 8 weeks of age) were randomly divided into three groups of six each. Group 1 was treated with 4g/kg body weight/day of the decoction for thirty days; rats in group 2 were treated with 6g/kg body weight/day for thirty days; Group 3 (control) was fed with distilled water (3 ml) for thirty days. On the 31st day, blood was collected by tail vein bleeding for assessment of effects on haematological parameters. The white blood cell (WBC) count and differential count (DC), red blood cell (RBC) count, packed cell volume (PCV) and haemoglobin (Hb) concentrations were determined according to the methods described by the International Committee for Standardization in Haematology (3). Using the above data, mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), and mean corpuscular haemoglobin concentration (MCHC) were calculated.

Effects on serum enzyme levels

Above experiment was continued for a further two months. At the end of the third month, rats were given ether anaesthesia and blood was collected by cardiac puncture. During this procedure the animal died and was subjected to an autopsy. Serum was analyzed for alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) according to methods described in commercially available reagent kits marketed by DMA Inc, USA.

Assessment of histological changes

At autopsy samples were taken from the heart, lungs, liver, kidneys, duodenum and stomach, and placed in 10% buffered formalin (sample volume: formalin volume=1:8) for histopathological studies. Histological changes (cell swelling, fatty change, granular and vacuolar degeneration and nuclear changes) were assessed by light microscopic examination of sections from the above organs stained with haematoxylin and eosin.

Effects on body weight gain, feed consumption ratio and body weight: liver weight ratio

Throughout the above experiment, the daily feed consumption and weekly body weight gain of each rat was recorded and average feed consumption and body weight gain was calculated. Before samples were taken from liver for histological examination, weight of each liver was recorded and body weight: liver weight ratio was calculated.

Effects on reproductive ability

Studies were done to determine effects of the decoction on reproductive ability of both

male and female rats. In female rats, effects on ovulatory activity and implantation were determined while in male rats effect on spermicidal activity was determined.

Effects on ovulatory activity

Wistar female rats (n=18, 14 to 16 weeks of age) with normal oestrus cycle (average 4-6 days of pro-oestrus, oestrus, metoestrus and dioestrus) were divided into three groups of six each. Group 1 and 2 were given decoction doses 4g/kg body weight/day and 6g/kg body weight/day respectively for 18 days. Group 3 was considered as the control group and 3ml of distilled water was given daily for 18 days. Vaginal smears were examined daily to check the stage of oestrus they were in. Persistent dioestrus, non-cyclic and lengthy oestrus cycles were considered as toxic signs. Different stages of the oestrus cycle were determined according to Soejarto *et al* (4).

Effects on implantation

Regularly cyclic female virgin rats (14 to 16 weeks of age) were mated. Presence of copulation plugs or sperms in the vaginal smear on the following morning were regarded as confirmation of mating. The day copulation plugs or sperms appeared was assumed the day one of pregnancy. Pregnant female rats (n=15) were randomly divided into three groups of five each. Groups 1 and 2 were given decoction doses 4g/kg body weight/day and 6g/kg body weight/day respectively from days 1 to 7 of pregnancy. Group 3 was given 3ml of distilled water from days 1 to 7. Autopsies were performed on the 10th day and the number of implantation sites, the number of live/dead fetuses and the number of *corpora lutea* of pregnancy were recorded (4).

Effects on spermicidal activity

Wistar male rats (n=18, 14 to 16 weeks of age) were randomly divided into three groups of six each. Groups 1 and 2 were given the decoction in doses of 4g/kg body weight/day and 6g/kg body weight/day respectively, for three months. Group 3 was considered as the control group and 3ml distilled water was given for three months. At the end of treatment, each male rat was mated individually with a female rat in oestrus (14 to 16 weeks of age). Presence of copulation plugs or sperms in the vaginal smear on the following morning were regarded as evidence of mating. After 18 to 21 days of gestation period, the number of mated female rats: number of unmated female rats, number of female rats who delivered pups, and the litter size of each female rat were recorded. All these parameters were considered as a reflection of the reproductive ability of the particular male rat, which mated each female rat (4).

Statistical analysis

Results are expressed as mean \pm standard error of mean (mean \pm SEM). Statistical analysis was done using the student's t test. $P < 0.05$ was considered as significant.

Results

LD₅₀ determination

No deaths were observed in the seven days following a single oral administration of several different doses of the decoction (60, 120, and 240 mg/kg body weight). Even the highest dose (240 mg/kg body weight) did not produce any apparent adverse effects. Therefore, these results did not allow the calculation of the LD₅₀.

Effects on haematological parameters

Table 1 summarizes the results obtained with respect to the effects of treatment with the decoction for 30 days on haematological parameters. No significant differences were observed between controls and treated rats concerning the parameters measured ($P>0.05$).

Serum enzyme levels

Table 2 summarizes the results found on analysis of ALT, AST and ALP after three months of decoction treatment. No significant differences were observed between controls and treated rats concerning the parameters measured ($P>0.05$).

Effects of the decoction on feed consumption, body weight gain, and body weight: liver weight ratio

Treatment with the decoction for three months did not alter average feed consumption, average body weight gain or body weight: liver weight ratio of the test animals. Results are shown in Table 3. No significant differences were seen between treated and the control group of rats ($P>0.05$).

Histopathology

No significant histological changes were observed in the heart, lung, liver, stomach, duodenum, and kidney of test animals when compared with the controls.

Table 1. A comparison of haematological variables of rats treated with the decoction for one month

Variable	Group 1 (decoction dose 4g/kg/day)	Group 2 (decoction dose 6g/kg/day)	Group 3 (distilled water control)
PCV %	52.8 ± 05	52.2 ± 0.4	53.3 ± 0.4
Hb (g/dl)	15.9 ± 0.4	15.5 ± 0.4	16.3 ± 0.6
RBC ($\times 10^6$ cells/mm ³)	8.5 ± 0.4	8.6 ± 0.4	8.4 ± 0.3
WBC ($\times 10^3$ cells/mm ³)	17.8 ± 0.3	14.8 ± 0.6	15.6 ± 1.2
Neutrophils ($\times 10^3$ cells/mm ³)	20.7 ± 1.2	19.5 ± 1.5	20.8 ± 0.3
Eosinophils ($\times 10^3$ cells/mm ³)	1.0 ± 0.3	2.0 ± 0.4	2.0 ± 0.3
Basophils ($\times 10^3$ cells/mm ³)	Nil	Nil	Nil
Monocytes ($\times 10^3$ cells/mm ³)	1.7 ± 0.2	2.5 ± 0.2	2.8 ± 0.5
Lymphocytes ($\times 10^3$ cells/mm ³)	77.2 ± 1.3	74.0 ± 2.2	86.8 ± 0.8
MCV (fl)	63.7 ± 2.0	60.8 ± 1.5	64.4 ± 2.4
MCH (pg)	18.9 ± 0.8	18.0 ± 0.3	19.6 ± 0.5
MCHC (g/dl)	31.0 ± 0.3	29.1 ± 0.8	30.3 ± 0.7

Data shown are the mean ± SEM of 10 determinants

None of the results were significantly different from control

Table 2. Effects of decoction on serum enzyme levels after three months of treatment

Group	ALT (IU/U)	AST (IU/L)	ALP (IU/L)
Group 1	23.6 ± 1.4	74.9 ± 6.0	67.8 ± 7.2
Group 2	18.4 ± 1.3	69.1 ± 6.3	73.0 ± 5.1
Group 3	19.4 ± 1.4	78.0 ± 6.9	84.0 ± 3.0

Data shown are the mean ± SEM of 10 determinants

Group 1: Decoction dose 4g/kg body weight/day

Group 2: Decoction dose 6g/kg body weight/day

Group 3: Control

None of the results were significantly different from control

Table 3. Effects of decoction on average feed consumption, average body weight gain and body weight: liver weight ratio

Group	Average feed consumption (g/day/rat)	Average body weight gain (g/day/rat)	Body weight: liver weight
Group 1	12.0 ± 1.8	1.6 ± 0.1	1.0 : 0.03
Group 2	11.5 ± 1.5	1.6 ± 0.1	1.0 : 0.03
Group 3	10.7 ± 1.4	1.6 ± 0.3	1.0 : 0.03

Data shown are the mean ± SEM of 10 determinants

Group 1: Decoction dose 4g/kg body weight/day

Group 2: Decoction dose 6g/kg body weight/day

Group 3: Control

None of the results were significantly different from control

Effects on reproductive ability**Anti-ovulatory activity**

As can be seen in Table 4, treatment with either of the two doses of decoction did not result in any alteration of the oestrus cycle, nor did the animals become persistently dioestrus.

Effects on implantation

Table 5 shows that oral administration of 4g/kg body weight/day of decoction did not significantly alter the number of pregnant animals, the number of non pregnant

animals, the number of implantation sites, the number of *corpora lutea* of pregnancy and the number of resorptions per total number of implantation sites ($P>0.05$).

Spermicidal activity

Results obtained are presented in Table 6. No significant differences were found after treatment with the decoction for three months in the number of pups delivered, or the litter size ($P>0.05$). The decoction therefore does not appear to have any significant effect on spermicidal activity of male rats.

Table 4. Effects of decoction on oestrus cycle

Group	Persistent dioestrus (number)	Regularly cyclic (number)	Duration of the oestrus (days)
Group 1	Nil	6.0±0.0	4.0 ±0.0
Group 2	Nil	6.0±0.0	4.0 ±0.0
Group 3	Nil	6.0±0.0	4.0 ±0.0

Data shown are the mean ± SEM of 10 determinants

Group 1: Decoction dose 4g/kg body weight/day

Group 2: Decoction dose 6g/kg body weight/day

Group 3: Control

None of the results were significantly different from control

Table 5. Effects of decoction on implantation

Variable	Group 1	Group 2	Group 3
Pregnancy rate	4/5 (80%)	3/5 (60%)	4/5 (80%)
Number of implantation sites	13.7 ± 1.5	13.6 ± 1.5	14.0 ± 1.4
Number of live foetuses	13.2 ± 2.1	13.3 ± 1.2	13.5 ± 1.0
Number of <i>corpora lutea</i> of pregnancy	14.8 ± 0.5	15.0 ± 1.7	14.2 ± 1.2
Number of resorptions/total implantation sites	2/53	1/40	2/56

Data shown are the mean ± SEM of 10 determinants

Group 1: Decoction dose 4g/kg body weight/day

Group 2: Decoction dose 6g/kg body weight/day

Group 3: Control

None of the results were significantly different from control

Table 6. Effects of decoction on spermicidal activity

Group	Number who delivered	Litter size
Group 1	6.0 ± 0.0	11.8 ± 0.4
Group 2	6.0 ± 0.0	12.0 ± 1.4
Group 3	6.0 ± 0.0	12.0 ± 0.6

Data shown are the mean ± SEM of 10 determinants

Group 1: Decoction dose 4g/kg body weight/day

Group 2: Decoction dose 6g/kg body weight/day

Group 3: Control

None of the results were significantly different from control

Discussion

For a herbal formulation to be of value clinically it should not only have the required pharmacological activities but must also be free of any serious adverse effects. Although a decoction of *N. sativa*, *H. indicus* and *S. glabra* has been in use for many years for the treatment of cancer patients, its possible adverse effects have not been evaluated in a scientifically controlled manner. Studies by other workers has shown that there are some medicinal plant extracts that contain toxic components, which have the potential to produce serious adverse reactions (5,6,7). Many of the plants used in the preparation of herbal teas, particularly *Senecio*, *Crotolaria* and *Heliotropium* species contain pyrrolizidine alkaloids that can be hepatotoxic (8). Similarly, some herbal extracts sold in capsule form in the market as stress relievers (eg. *Kalms*, *Neurolex*) have been reported to produce severe liver damage in people who consume these extracts (6). It is therefore very important to carry out scientifically controlled studies on these remedies to ensure that they are safe and will not produce any serious adverse effects.

According to Arsecularatne *et al* (9) and Tennekoon *et al* (10), *H. indicus* and *N. sativa* respectively, contain hepatotoxic components. However, in the present study, despite the extensive toxicity investigations, the tested decoction did not produce any significant toxicological effects when administered in the doses recommended by the traditional medical practitioners. A recent study also showed that *N. sativa* does not have any toxicological properties (11). The explanation for the discrepancy could be that the dose of *N. sativa* used by Tennekoon *et al* was about four times higher than that used as the highest dose of decoction in the present study (2g/kg/day). Arsecularatne *et al* 1985 incorporated

powdered *H. indicus* directly into the rat feed whereas in the present investigation, researchers fed the decoction containing *H. indicus* orally. Whether these two types of feeding have any difference regarding the constituents is unclear.

According to Nagaratnam *et al* (12), alkaloids in various decoctions consumed by people in Sri Lanka may cause liver damage, leading to hepatocellular carcinoma. However, the decoction used in the present study protected against chemically induced hepatocarcinogenesis (1) and did not cause any liver damage, as assessed by lack of alterations in the serum enzyme levels AST, ALT and ALP and liver histopathology, even after three months of continuous treatment. Whether the difference in effects is due to species difference or due to difference in the chemical components of the decoction used in the study requires investigation.

Zaoui *et al* (13) had reported that fixed oil of *N. sativa* could cause leukocyte counts to decrease and PCV and haemoglobin levels to increase in rats. However, present study with decoction containing *H. indicus* and *S. glabra* in addition to *N. sativa*, revealed that none of the haematological parameters that were tested were altered even after three months of continuous treatment. The same investigation by Zaoui *et al* also showed that fixed oil of *N. sativa* caused a slowing of body weight gain in rats. However, in the present study no changes in body weight gain was observed during the experimental period. Whether these differences are due to protective effects of components in *H. indicus* and *S. glabra* is not clear.

It is well known that several commonly used plant based drugs have adverse effects on fertility, as well as teratogenic effects (14).

Morphine and caffeine given to male rats before mating decreases the number of pups and frequently lead to death (15). In the present study where the effects of the decoction on ovulatory activity, implantation and spermicidal activity were investigated, no alterations were found in the length of oestrus cycle, the interval between cycles, gestation period and number of pups per litter or any other parameter investigated.

The general well being of the animals also did not change. All the animals remained in good health throughout the experimental period. This study has shown that this decoction did not produce any significant adverse effects on body systems in rats and mice.

Acknowledgements

We thankfully acknowledge the research grant from National Science Foundation, Sri Lanka and Ayurveda Dr. N. Jayathilake for providing the recipe for preparation of the decoction. Authors also wish to thank Dr. S. Jayasekera and Mr. Sarath Sisira Kumara, Animal Centre, MRI, Colombo and Ms. Sujatha Ramadasa, Department of Pathology, Faculty of Medicine, University of Peradeniya, for the support provided.

References

- Iddamaldeniya S.S., Wickremasinghe N., Thabrew M.I., Ratnatunge N., Thammitiyagodage M.G. Protection against diethylnitrosamine-induced hepatocarcinogenesis by an indigenous medicine comprised of *Nigella sativa*, *Hemidesmus indicus*, and *Smilax glabra*: a preliminary study. *Journal of Carcinogenesis* 2003; 2: 6.
- Sabourdy M.A. Breeding and care of laboratory animals, Volume 1, pp. 18-19. WHO 1988, Health Laboratory Technology Unit, Geneva, Switzerland.
- International Committee for Standardization in Haematology. Recommendations for reference method for determination of centrifugation of packed cell volume of blood. *Journal of Clinical Pathology* 1980; 33: 1.
- Soejarto D.D., Bingel A.S., Slaytor M., Farnsworth N.R. Fertility regulating agents from plants. *Bulletin of the World Health Organization* 1978; 56: 343-352.
- Penn R.G.C. Adverse reactions to herbal medicines. *Australian Prescriber* 1988; 2: 15-18.
- McGregor F.B., Abernethy V.E., Dahabra S., Cobden I., Hades P.C. Hepatotoxicity of herbal remedies. *British Medical Journal* 1989; 299: 1156-1158.
- Maruo V.M., Soares M.R., Bernardi M.M., Spinosa H.S. Embryotoxic effects of *Solanum lycocarpum* St. Hill fruit consumption during preimplantation and organogenesis in rats. *Neurotoxicology and Teratology* 2003; 25: 627-631.
- Arseculeratna S.N., Gunathilake A.A.L., Panabokke R.G. Studies on medicinal plants of Sri Lanka: occurrence of pyrrolizidine alkaloids and hepatotoxic properties in some traditional medicinal herbs. *Journal of Ethnopharmacology* 1981; 4: 159-177.
- Arseculeratna S.N., Gunathilake A.A.L., Panabokke R.G. Studies on medicinal

- plants of Sri Lanka Part 14: Toxicity of some traditional medicinal herbs. *Journal of Ethnopharmacology* 1985; 13: 323-335.
10. Tennekoon K.H., Thyaparam J., Kurukulasooriya A.P., Karunanayake E.H. Possible hepatotoxicity of *Nigella sativa* and *Dregea volubilis* leaves. *Journal of Ethnopharmacology* 1990; 31: 283-289.
 11. Ali B.H., Blunden G. Pharmacological and toxicological properties of *Nigella sativa*. *Phytotherapy Research* 2003; 17: 299-305.
 12. Nagaratnam N., Ramachandran V., Jiffrey A.J., Bhuvanendran N. Primary liver cancer in Sri Lanka. *Journal of Tropical Medicine and Hygiene* 1984; 87: 185-187.
 13. Zaoui A., Cherrah Y., Mağassini N., Alaoui K., Amarouch H., Hassar M. Acute and chronic toxicity of *Nigella sativa* fixed oil. *Phytomedicine* 2002; 9: 69-74.
 14. Oliveira M.G.M., Monteiro M.G., Macaubas C., Barbosa V.P., Carlini E.A. Pharmacologic and toxicologic effects of two *Maytenus* species in laboratory animals. *Journal of Ethnopharmacology* 1991; 34: 29-41.
 15. Joffe J.M. Influence of the drug exposure of the father on perinatal outcome. *Clinics in Perinatology* 1979; 67: 21-36.